

## Chapter 15

# Biological N Inputs

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**TABLE S15.1** Examples of genera of diazotrophic bacteria arranged by mode of energy generation and the oxygen sensitivity of their diazotrophy

Energy source	Sensitivity of N <sub>2</sub> fixation to oxygen	Examples (Genera)
Heterotrophic	Aerobic diazotrophs	<i>Azotobacter</i> , <i>Gluconacetobacter</i>
	Microaerophilic diazotrophs	<i>Azospirillum</i> , <i>Herbaspirillum</i> , <i>Methylococcus</i>
	Facultatively aerobic diazotrophs	<i>Klebsiella</i> , <i>Paenibacillus</i> <i>Enterobacter</i>
	Obligately anaerobic diazotrophs	<i>Clostridium</i> , <i>Desulfovibrio</i> , <i>Methanosarcina</i>
Phototrophic	Aerobic diazotrophs	<i>Anabaena</i> , <i>Nostoc</i>
	(primarily filamentous heterocyst-forming cyanobacteria)	
	Microaerophilic diazotrophs (filamentous nonheterocystous cyanobacteria)	<i>Lyngbya</i> , <i>Oscillatoria</i>
	Facultatively aerobic diazotrophs (purple nonsulfur bacteria)	<i>Rhodobacter</i>
	Obligately anaerobic diazotrophs (purple sulfur bacteria)	<i>Chromatium</i>

Adapted from Young, 1992. With kind permission of Springer Science + Business Media B.V. From Soil Microbiology, Ecology, and Biochemistry, 3<sup>rd</sup> Edition, Paul, E.A, Biological N inputs (2007). pp. 365–387.

**TABLE S15.2** Examples of Nitrogen fixing (*nif*) genes, their products and functions

Gene	Product and function
<i>nifH</i>	Nitrogenase reductase subunit; binds two molecules of MgATP and reduces nitrogenase by single electron transfers
<i>nifD</i>	Nitrogenase protein alpha subunit of dinitrogenase
<i>nifK</i>	MoFe protein beta subunit of dinitrogenase
<i>nifF</i>	Flavodoxin, reductant of nitrogenase reductase
<i>nifJ</i>	Pyruvate-flavodoxin oxidoreductase; couples oxidation of pyruvate to reduction of flavodoxin
<i>nifS</i>	Pyridoxal-dependent cysteine desulfurase: required for synthesis of <i>nifB</i> -FeMo cofactor intermediate
<i>nifU</i>	Complements <i>nifS</i> . Required for assembly of <i>nifB</i> -FeMo cofactor intermediate-mobilization of Fe and S for metallocluster assembly and synthesis of active enzyme.
<i>nifV</i>	Homocitrate synthase; organic component of FeMo cofactor
<i>nifN</i>	Subunit of $nifN_2E_2$ which provides a transient site for the assembly of FeMo cofactor.
<i>nifE</i>	Subunit of $nifN_2E_2$
<i>nifB</i>	FeMo cofactor precursor biosynthesis
<i>nifQ</i>	Early step in FeMo cofactor biosynthesis
<i>nifX</i>	Intermediate carrier in FeMo cofactor biosynthesis
<i>nifY</i>	Intermediate carrier in FeMo cofactor biosynthesis
<i>nifW</i>	Required for synthesis of fully active dinitrogenase
<i>nifZ</i>	Required for synthesis of fully active dinitrogenase
<i>nifA</i>	Positive regulatory protein
<i>nifL</i>	Negative regulatory protein
<i>nifT</i>	Unknown function
<i>naY</i> <i>clpX</i> <i>rnf</i> ABCDEGH	Probably an intermediate carrier in FeMo cofactor biosynthesis Involved in controlling proteolytic cleavage of FeMoCo cofactor biosynthesis proteins <i>NifE</i> <i>NifN</i> <i>NifB</i> Required for maturation of an active nitrogenase reductase

Data from Dean and Jacobsen (1992); Setubal et al. (2009); Dos Santos and Dean, (2011).

**TABLE S15.3** Examples of genera and species of the root nodule bacteria of legumes and their hosts

Genera and representative species
<p><b>Azorhizobium</b> (2 species)  <i>A. caulinodans</i> (<i>Sesbania</i>)</p>
<p><b>Bradyrhizobium</b> (~13 species)  <i>B. arachidis</i> (peanut)  <i>B. canariense</i> (genistoid legumes)  <i>B. cytisus</i> (<i>Cytisus</i>, broom)  <i>B. elkanii</i> (soybean)  <i>B. japonicum</i> (soybean)  <i>B. liaoningense</i> (soybean)  <i>B. sp.</i> (<i>Vigna</i>, <i>Lupinus</i>, etc.)<sup>a</sup></p>
<p><b>Mesorhizobium</b> (~20 species)  <i>M. amorphae</i> (<i>Amorpha</i>)  <i>M. chacoense</i> (<i>Prosopis</i>, mesquite)  <i>M. ciceri</i> (<i>Cicer</i>, chickpea)  <i>M. huakii</i> (<i>Astragalus</i>, milkvetch)  <i>M. loti</i> (<i>Lotus</i>, trefoil)  <i>M. mediterraneum</i> (<i>Cicer</i>, chickpea)  <i>M. plurifarum</i> (tropical trees)</p>
<p><b>Rhizobium</b> (~30 species)  <i>R. etli</i> (<i>Phaseolus</i>, bean)  <i>R. galegae</i> (<i>Galega</i>)  <i>R. gallicum</i> (<i>Phaseolus</i>, bean)  <i>R. giardinii</i> (bean)  <i>R. huakuii</i> (<i>Astragalus</i>)  <i>R. huautlense</i> (<i>Sesbania</i>)  <i>R. indigofera</i> (<i>Indigofera</i>)  <i>R. leguminosarum</i> (three biovars nodulate: (i) clovers, (ii) peas, lentils, and vetch, (iii) bean)  <i>R. mongolense</i> (<i>Medicago</i>)  <i>R. phaseoli</i> (beans)  <i>R. tropici</i> (<i>Phaseolus</i>, <i>Leucaena</i>, bean)</p>
<p><b>Ensifer</b>, formerly <b>Sinorhizobium</b> (~10 species)  <i>E. americanum</i> (<i>Acacia</i>)  <i>E. arboris</i> (tree legumes)  <i>E. fredii</i> (soybean)  <i>E. kostiense</i> (tree legumes)  <i>E. kummerowiae</i> (<i>Kummerowia</i>)  <i>E. medicae</i> (annual medics)  <i>E. meliloti</i> (alfalfa)  <i>E. morelense</i> (<i>Leucaena</i>)  <i>E. saheli</i> (<i>Sesbania</i>)  <i>E. teranga</i> (<i>Sesbania</i>)</p>

**TABLE S15.3** Examples of genera and species of the root nodule bacteria of legumes and their hosts—Cont'd***Burkholderia*** (~10 species)*B. phymatum* (nodulates *Mimosa* spp.)*B. tuberum* (nodulates *Cyclopia*, *Macroptilium* and *P. vulgaris*, not *Mimosa* spp.)

<sup>a</sup>Although many legumes are nodulated by bacteria of the *Bradyrhizobium* genus, the latter have not received official species designation. They are referred to by the name of the legume host from which they were isolated, (e.g., *Bradyrhizobium* [Lupinus]).

There are several other genera of alpha-proteobacteria with legume-nodulating properties including *Devosia*, *Cupravidus*, *Ochrobactrum*, *Methylobacterium*, and *Phyllobacterium*.

Taken from the website of the ICSP subcommittee on the taxonomy of *Rhizobium* and *Agrobacterium*, <http://edzna.ccg.unam.mx/rhizobial-taxonomy/>

